

STUDIES ON INDIAN MEDICINAL PLANTS—PART LXXV¹

NISHINDASIDE, A NOVEL IRIDOID GLYCOSIDE FROM VITEX NEGUNDO

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Abstract - Two new glucosidic iridoids, designated as nishindaside ($2a$) and negundoside ($3a$), have been isolated from the leaves of Vitex negundo (Verbenaceae). The absence of a C(3)-C(4) double bond and the presence of an equatorial 3-methoxy group in $2a$, the 3-methoxy-3,4-dihydro derivative of agnuside ($1a$), were established from 1H and ^{13}C NMR analyses, corroborated by mass spectral fragmentation of the hexaacetate ($2b$). Negundoside ($3a$) could be concluded to be the 2'-*p*-hydroxy benzoyl derivative of mussaenosidic acid from the results of alkaline hydrolysis and the shifts in the signals of C-1', C-2' and C-3' in its ^{13}C NMR spectrum.

Vitex negundo L. (local name Nishinda), belonging to the family Verbenaceae, is reputed to have antiinflammatory and tiarthritic properties in the Indian stem of medicine². Earlier workers reported³⁻¹¹ the isolation of a variety of constituents including the iridoid agnuside ($1a$). Re-examination of the ethanolic extract of its leaves led us to two other iridoid glycosides, designated as nishindaside ($2a$) and negundoside ($3a$). The structure determination of $2a$ which is discussed in the present paper.

Nishindaside, amorphous, $[M]_D^{25} - 3.5^\circ$, appeared to be an iridoid glycoside from colour reactions and the

similarity of its IR spectrum with that of $1a$. Absence of the characteristic band at $1645-1650\text{ cm}^{-1}$, however, ruled out the possibility of a $-O-C(3)=C(4)-$ moiety, usually present in the iridoids.

Though the M^+ ion could not be detected, significant peaks at m/z 138, 121, 93 and 65 in its mass spectrum provided evidence for the presence of a monohydroxy benzoyl group in the molecule. That the compound is in fact an ester of *p*-hydroxy benzoic acid was evident from a symmetrical double doublet at δ 7.86 and 6.86 (J 8 Hz) in its 1H NMR spectrum (Table 1) which also provided additional structural information as in the sequel.

A 3H singlet at δ 3.40 indicated a methoxy group attached to the iridoid nucleus. The doublets at δ 4.96 (J 6Hz) and 4.52 (J 8 Hz) could be assigned to two acetal protons, the chemical shift and the coupling constant of the latter being commensurate with a β -glycoside. Furthermore, a broad singlet at δ 5.76 suggested a trisubstituted double bond in the cyclopentane ring. The absence of any other olefinic proton signals, however, corroborated the absence of the C(3)-C(4) double bond.

Hydrogenation of nishindaside with Adams catalyst afforded **4**, the ^1H NMR spectrum of which showed no signal for the vinylic proton and the aromatic moiety. Instead, a 3H doublet was observed at δ 1.08 (J 7 Hz) for a methyl group obviously formed by hydrogenolysis of an allylic ester. The above facts strongly supported the location of the trisubstituted double bond between C(7) and C(8) and the attachment of the aromatic ester group to C(10) of the iridoid nucleus.

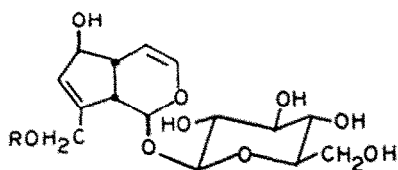
On treatment with $\text{Ac}_2\text{O}/\text{Py}$ at room temperature, nishindaside afforded a crystalline hexaacetate (**2b**). Its high field (400 MHz) ^1H NMR spectrum exhibited five singlets in the region δ 2.0-2.1 ppm for aliphatic acetates and a more deshielded signal at δ 2.31 assignable to an aromatic one. Of the aliphatic acetoxy groups, four could be located in the sugar residue which was identified as glucose from the observed all diaxial coupling constants for 2'-H, 3'-H and 4'-H. That the remaining one was allylic to the double bond became apparent from the 1H multiplet at δ 5.60. In addition, the existence of an equatorial methoxy group at C-3 could be deduced from the double doublet at δ 4.75 ($J_{\text{AX}} + J_{\text{BX}} = 13$ Hz) assigned to the axial C(3) proton.

Like the parent compound, the hexaacetate (**2b**) also did not show the molecular ion in the mass spectrum (CI) but the primary fragment at m/z 691 could easily be rationalised by assuming the loss of a molecule of acetic acid from $\text{[M} + \text{H}]^+$. Further elimination of 32 mass units, compatible with the expulsion of methanol, secured the presence of the methoxy group in the molecule. The spectrum exhibited characteristic peaks for the sugar and the aromatic ester moieties as well.

All the above evidences therefore led to structure **2a** for nishindaside. It was further corroborated by ^{13}C NMR spectrometry. For this purpose, the spectrum of **1a** had to be examined first. It displayed separate signals for all the carbons (except for the chemically equivalent pairs 2", 6" and 3", 5"), The individual assignments (Table 2), based on the splittings in the SFORD spectrum and chemical shift considerations, are in very good agreement with the reported data for aucubin (**1b**)¹² and melampyroside (**1c**)¹³.

The chemical shifts of the carbons of **2a** were found to be very close to those of **1a** and the disappearance of signals at δ 141.6 and 105.4 in the spectrum of **2a** was in conformity with the absence of the 3,4-unsaturation. Additional signals observed at δ 100.0 and 30.0 ppm could however be assigned to C-3 and C-4, the more deshielded one being assigned to C-3 carrying a methoxy group,

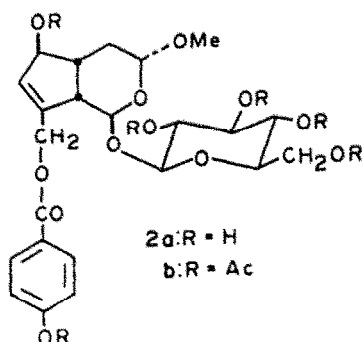
The other compound, negundoside (**2a**), m.p. 160-64 $^\circ$, $\text{[}\alpha\text{]}_D^{25} -130.0^\circ$, could be inferred to be an $\alpha\beta$ -unsaturated 4-carboxy iridoid carrying a *p*-disubstituted aromatic ring from the characteristic ^1H NMR signals.



1a: R = COC₆H₄OH (p)

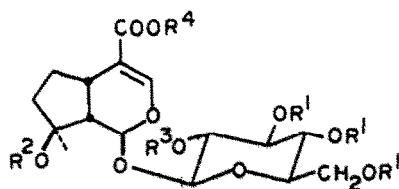
b: R = H

c: R = COC₆H₅



2a: R = H

b: R = Ac



3a: R¹ = R² = R⁴ = H, R³ = COC₆H₄OH (p)

b: R¹ = R² = R³ = R⁴ = H

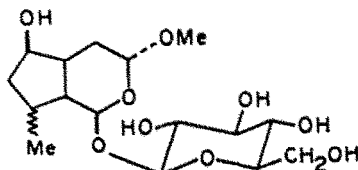
c: R¹ = Ac, R² = R⁴ = H, R³ = COC₆H₄OH (p)

d: R¹ = Ac, R² = R⁴ = H, R³ = COC₆H₄OAc (p)

e: R¹ = Ac, R² = H, R³ = COC₆H₄OMe (p),

R⁴ = Me

f: R¹ = R² = Ac, R³ = COC₆H₄OAc (p), R⁴ = H



4

Treatment of \mathfrak{A}_a with methanolic alkali at room temperature afforded methyl-p-hydroxy benzoate and muscadinic acid (\mathfrak{A}_b)¹⁴. The linkage of the aromatic ester moiety at position 2' is established from a downfield shift of C-2' and shielding of C-1' and C-3' signals in the ¹³C NMR spectra of \mathfrak{A}_a compared to \mathfrak{A}_b .

Since the structure of probably identical compound has recently been reported¹⁵, only the additional information related to the structure identification is presented herein.

Acetylation of \mathfrak{A}_a with Ac₂O/Py at room temperature in our hands yielded triacetate (\mathfrak{A}_c) besides the tetraacetate (\mathfrak{A}_d)¹⁵. The former furnished an ethyl derivative (\mathfrak{A}_e) which exhibited two 3H singlets at δ 3.84 and 3.40 in the ¹H NMR spectrum corroborating thereby the presence of a phenolic and carboxylic group in negundoside (\mathfrak{A}_a).

On the other hand, the acetylation of \mathfrak{A}_a at higher temperature led to a pentaacetate (\mathfrak{A}_f); The downfield shift of the methyl, 1-H and 9-H signals in its ¹H NMR spectrum over those of \mathfrak{A}_a adduced evidence for the β -oriented tertiary hydroxyl group¹⁶ in negundoside (\mathfrak{A}_a). The same conclusion could also be arrived at from the chemical shift (δ 52.2) of C-9 in the ¹³C NMR spectrum of \mathfrak{A}_a [cf.¹⁷ δ 50 \pm 2 ppm for C₉-OH (β) and 44.5 \pm 1 ppm for C₉-OH (α)].

Nishindaside is a unique 3,4-dihydro C₉-iridoid carrying a 3-methoxy group hitherto not encountered in nature, while negundoside belongs to the rare 2'-esterified iridoids, the only other compounds being reported very recently from Scrophulariaceae¹⁸ and Rubiaceae¹⁹.

Table 1
¹H NMR Spectral data of the iridoid glycosides^{a, b}

Position	1a	2a	3a	4a	5a	6a	7a	8a	9a
1		4.96d (6)	5.21d (3)	5.40d (4)	5.36d (4)				5.80brs
3	6.36dd (2, 7)		4.75dd (6, 7)	7.04s	7.28s	7.12s	7.04s	7.10s	7.10s
4	5.10dd (4, 7)								
5	2.90m		2.65m	2.70m			2.80m	2.90m	2.80m
6	4.38m		5.60m						
7	5.80brs	5.76brs	5.86brs						
9	2.90m		3.15brd (9)	2.06dd (3, 9)	2.10dd (4, 9)	2.26dd (4, 11)		2.28dd (4, 10)	2.65brd (8)
10	4.90brs	4.84brs	4.92 4.98 ^c	1.14s	1.20s	1.26s	1.24s	1.28s	1.44s
1'	4.54d (8)	4.52d (8)	4.88d (8)	4.85d (8)	4.56d (8)	5.00d (8)	4.96d (8)	5.00d (8)	4.98d (8)
2'			5.02dd (8, 10)	4.75t (8)		5.10- 5.50m	5.10- 5.50m	5.10- 5.50m	5.10- 5.60m
3'			5.08t (10)			5.10- 5.50m	5.10- 5.50m	5.10- 5.50m	5.10- 5.60m
4'			5.28t (10)			5.10- 5.50m	5.10- 5.50m	5.10- 5.50m	5.10- 5.60m
5'			3.70ddd (2, 5, 10)				3.85brd (10)		3.86brd (10)
6'	3.7m		4.05dd (2, 13) 4.19dd (5, 13)			4.26m	4.26m	4.26m	4.20dd (2, 12) 4.46dd (5, 12)
2'', 6''	7.90d (8)	7.86d (8)	8.10d (9)	7.80d (8)		7.80d (9)	8.01d (8)	7.90d (8)	8.01d (8)
3'', 5''	6.90d (8)	6.86d (8)	7.20d (9)	6.85d (8)		6.80d (9)	7.11d (8)	6.90d (8)	7.10d (8)
OCH ₃		3.40s	3.41s					3.4s 3.84s	
OCOCH ₃			2.00- 2.10, 5xs 2.31s			1.95s 2.06s 2.12s	1.96s 2.04s 2.12s 2.36s	1.90s 2.06s 2.10s	1.95s 1.98s 2.04s 2.12s 2.36s

^aδ (ppm) downfield from TMS, J values in parentheses.

^bSolvents: 1a, 2a, 3a, 4a in DMSO-d₆; 5a, 6a, 7a, 8a in CDCl₃ and 9a in CDCl₃-CD₃OD.

^cInner limbs of an ABq, outer limbs merged with other signals.

Table 2
 ^{13}C NMR chemical shifts in CD_3OD

Carbon	1a	2a	3a	3b
	97.9	98.4	94.9	95.1
	141.6	100.0	151.0	151.6
	105.4	30.2	113.5	*
	46.1	44.6	30.9	31.8
	82.7	80.8	30.0	30.5
	132.3	131.4	41.2	40.7
	142.7	142.5	79.7	80.4
	48.4	49.2	52.2	52.1
	63.5	63.0	24.3	24.5
	-	-	169.8	*
	100.1	100.0	97.6	99.6
	74.7	74.6	75.8	74.5
	78.0	77.9	74.7	78.0
	71.3	71.2	71.6	71.5
	77.8	77.9	78.1	77.7
	62.6	62.6	62.6	62.7
	121.9	121.8	122.0	-
,6"	132.7	132.7	132.7	-
,5"	116.2	116.1	116.0	-
	163.5	163.3	163.0	-
H_3	-	56.1	-	-
	167.7	167.6	167.1	-

The quaternary carbon signals not observed probably due to high relaxation time.

EXPERIMENTAL

All melting points are uncorrected. IR spectra were recorded on a Perkin-Elmer infrared spectrophotometer (model 577) and mass spectra (EI) on a Hitachi RU-6L instrument. ^1H and ^{13}C NMR spectra were mostly measured on a Jeol FX-100 FT IR spectrometer using TMS as internal standard.

Isolation of the iridoid glycosides. Dried leaves (5 kg) of *Vitex negundo* were successively extracted in a Soxhlet with petrol, CHCl_3 and MeOH. The ethanolic extract was concentrated to a syrup, triturated with H_2O and the aqueous layer exhaustively extracted with $n\text{-BuOH}$. The residue (400 g) obtained after complete evaporation of BuOH was

chromatographed repeatedly over silica gel, eluting with CHCl_3 with increasing MeOH content. The first few fractions eluting with 8% MeOH/ CHCl_3 were pooled; evaporation gave pure agnuside (1a). The subsequent fractions on evaporation gave a mixture of 1a and 2a from which 2a was purified by prep. TLC. The 10% MeOH/ CHCl_3 eluents afforded compound 3a.

Agnuside (1a). Recrystallisation from MeOH- CHCl_3 yielded agnuside (3 g), m.p. 148-52°, IR (nujol): ν_{max} 3400, 1708, 1640, 1590, 1455 cm^{-1} .

Nishindaside (2a). Obtained as an amorphous powder (150 mg), $\Delta\alpha_D^{25} -83.5^\circ$ (MeOH, c 0.974), IR (KBr): ν_{max} 3400 (OH), 1700 (CO), 1605, 1590, 1450 (aromatic) cm^{-1} .

Hexa-O-acetyl derivative of 2a.

Compound 2a (25 mg) was treated with dry pyridine (0.1 ml) and Ac_2O (0.1 ml) at room temp and left overnight. After addition of 0.1 ml MeOH, the solution was left for 20 min, then evaporated to give 3b (22 mg), crystallised from CHCl_3 -petrol as needles, m.p. 161-62°, $\Delta\alpha_D^{25} -71.4^\circ$ (CHCl_3 , c 0.68), IR (KBr): ν_{max} 1740 cm^{-1} ; CI-MS (Isobutane): m/z (rel.int.%) 691(1), 659(2), 331(97), 271(95), 229(37), 181(97), 169(92), 162(98), 139(97), 121(80), 109(39), 57(100).

Hydrogenation of 2a. Compound 2a (20 mg) was hydrogenated using PtO_2 (2 mg) in 4 ml EtOH until H_2 uptake ceased. After usual work up and purification, 4 (15 mg) was obtained as an amorphous material.

Negundoside (3a). Crystallised from MeOH- CHCl_3 as white needles (1g), m.p. 160-64°, $\Delta\alpha_D^{25} -130^\circ$ (MeOH, c 0.122), IR (nujol): ν_{max} 3400 (OH), 1710, 1690 (CO), 1640, 1605, 1590 and 1455 cm^{-1} .

Acetylation of 3a at room temp.

Compound 3a (100 mg) was acetylated with pyridine (0.5 ml) and Ac_2O (0.5 ml) at room temp overnight. Usual work up followed by purification through column chromatography yielded the products 3c (40 mg) and 3d (35 mg).

Tri-O-acetyl derivative of 3a.

Compound 3c, amorphous, $\Delta\alpha_D^{25} -91.6^\circ$ (MeOH, c 0.936), IR (nujol): ν_{max} 3350, 1740, 1710, 1620 and 1230 (OCOCH_3) cm^{-1} .

Tetra-O-acetyl derivative of 3a.
 3d, m.p. 118-20°, $\angle \alpha_D^{25}$ -96.6° (CHCl₃, c 0.17), IR (CHCl₃): ν_{\max} 1740, 1710 (CO), 1635, 1600 and 1240 (OCOCH₃) cm⁻¹.

Penta-O-acetyl derivative of 3a.
 Compound 3a (60 mg) was acetylated with pyridine (0.3 ml) and Ac₂O (0.3 ml) at 100° for one hr and worked up in the usual way. Purification by column chromatography yielded two products 3d (20 mg) and 3e (30 mg). Compound 3e, an amorphous powder, $\angle \alpha_D^{25}$ -48.4° (CHCl₃, c 0.64); IR (CHCl₃): ν_{\max} 1745, 1720, 1695, 1630, 1600, 1230 (OCOCH₃) cm⁻¹.

Methylation of 3c. Treatment of 3c (20 mg) with CH₂N₂ overnight afforded the amorphous compound 3e (15 mg), $\angle \alpha_D^{25}$ -57.6° (CHCl₃, c 0.26), IR (nujol): ν_{\max} 3400, 1740 (CO), 1690, 1635, 1600 and 1230 (OCOCH₃) cm⁻¹.

Hydrolysis of 3a. Compound 3a (50 mg) was hydrolysed by keeping it overnight with 0.1N NaOH (1 ml) and then neutralising by passing CO₂. Purification by column chromatography afforded methyl-p-hydroxy benzoate and compound 3b (30 mg).

Mussaenoside tetraacetate and pentaacetate. Compound 3b (50 mg) was methylated with CH₂N₂ overnight and the crude material acetylated with Ac₂O/Py at 100°. Usual work up followed by chromatographic purification yielded mussaenoside tetraacetate (20 mg) and mussaenoside pentaacetate (15 mg), identified by direct comparison (co TLC, IR, ¹H NMR) with authentic specimen.

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